

MLH1 Hypermethylation Analysis, Blood

Test ID: MLHPB

Explanation: On the effective date, the Specimen Stability and Specimen Required will be updated for this assay. *Any anticoagulant* for whole blood specimens will no longer be accepted and additional specimen types will be added.

Current Specimen Stability			
Specimen Type	Temperature	Time	Special Container
Varies	Ambient (preferred)		
	Frozen		
	Refrigerated		

New Specimen Stability			
Specimen Type	Temperature	Time	Special Container
Varies	Varies		

Current Specimen Required
<p>Patient Preparation: A previous bone marrow transplant from an allogenic donor will interfere with testing. Call 800-533-1710 for instructions for testing patients who have received a bone marrow transplant.</p> <p>Container/Tube: Preferred: Lavender top (EDTA) or yellow top (ACD)</p> <p>Acceptable: Any anticoagulant</p> <p>Specimen Volume: 3 mL</p> <p>Collection Instructions:</p> <ol style="list-style-type: none"> Invert several times to mix blood. Send whole blood specimen in original tube. Do not aliquot.

New Specimen Required
<p>Patient Preparation: A previous bone marrow transplant from an allogenic donor will interfere with testing. Call 800-533-1710 for instructions for testing patients who have received a bone marrow transplant.</p> <p>Submit only 1 of the following specimens:</p> <p>Specimen Type: Whole blood Container/Tube: Lavender top (EDTA) or yellow top (ACD) Specimen Volume: 3 mL</p> <p>Collection Instructions:</p> <ol style="list-style-type: none"> Invert several times to mix blood. Send whole blood specimen in original tube. Do not aliquot. <p>Specimen Stability Information: Ambient (preferred) 4 days/Refrigerated 4 days/Frozen 4 days</p> <p>Additional Information:</p> <ol style="list-style-type: none"> Specimens are preferred to be received within 4 days of collection. Extraction will be attempted for specimens received after 4 days, and DNA yield will be evaluated to determine if testing may proceed. To ensure minimum volume and concentration of DNA is met, the requested volume must be submitted. Testing may be canceled if DNA requirements are inadequate.

Specimen Type: Cord blood**Container/Tube:**

Preferred: Lavender top (EDTA) or yellow top (ACD)

Specimen Volume: 3 mL

Collection Instructions:

1. Invert several times to mix blood.
2. Send cord blood specimen in original tube. Do not aliquot.

Specimen Stability Information: Ambient (preferred) 4 days/Refrigerated 4 days/Frozen 4 days

Additional Information:

1. Specimens are preferred to be received within 4 days of collection. Extraction will be attempted for specimens received after 4 days, and DNA yield will be evaluated to determine if testing may proceed.
2. To ensure minimum volume and concentration of DNA is met, the requested volume must be submitted. Testing may be canceled if DNA requirements are inadequate.
3. While a properly collected cord blood sample may not be at risk for maternal cell contamination, unanticipated complications may occur during collection. Therefore, maternal cell contamination studies are recommended to ensure the test results reflect that of the patient tested and are available at an additional charge. Order MATCC / Maternal Cell Contamination, Molecular Analysis, Varies on the maternal specimen.

Specimen Type: Extracted DNA**Container/Tube:**

Preferred: Screw Cap Micro Tube, 2 mL with skirted conical base

Acceptable: Matrix tube, 1 mL

Collection Instructions:

1. The preferred volume is at least 100 mcL at a concentration of 75 ng/mcL.
2. Include concentration and volume on tube.

Specimen Stability Information: Frozen (preferred) 1 year/Ambient/Refrigerated

Additional Information: DNA must be extracted in a CLIA-certified laboratory or equivalent and must be extracted from a specimen type listed as acceptable for this test (including applicable anticoagulants). Our laboratory has experience with Chemagic, Puregene, Autopure, MagnaPure, and EZ1 extraction platforms and cannot guarantee that all extraction methods are compatible with this test. If testing fails, one repeat will be attempted, and if unsuccessful, the test will be reported as failed and a charge will be applied. If applicable, specific gene regions that were unable to be interrogated due to DNA quality will be noted in the report.

Questions

Contact Michelle Rath, Laboratory Resource Coordinator at 800-533-1710.